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### (54) **Polymorphisms in the human organic anion transporter C (OATP-C) gene**

Polymorphismen im Gen für den humanen organischen Anionentransporter C (OATP-C)

Polymorphismes dans le gène humain du transporteur d'anion organique (OATP-C)

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(56) References cited:  
**WO-A-00/08157 WO-A-99/06046**

- **TAMAI I ET AL: "MOLECULAR IDENTIFICATION AND CHARACTERIZATION OF NOVEL MEMBERS OF THE HUMAN ORGANIC ANION TRANSPORTER (OATP) FAMILY" BIOCHIMICA ET BIOPHYSICA ACTA, AMSTERDAM, NL, vol. 273, no. 1, 2000, pages 251-260, XP000941538 ISSN: 0006-3002 & DATABASE EMBL [Online] EBI; 8 June 1999 (1999-06-08) TAMAI I ET AL.: "Homo sapiens mRNA for organic anion transporter OATP-C" retrieved from [HTTP://WWW.EBI.AC.UK/CGI-BIN/EMBLFETCH](http://www.ebi.ac.uk/cgi-bin/emblfetch) Database accession no. AB026257**
- **DATABASE EMBL [Online] EBI; 2 February 2000 (2000-02-02) MUZNY DM ET AL.: "Homo sapiens 12p BAC RP11-12505" retrieved from [HTTP://WWW.EBI.AC.UK/CGI-BIN/EMBLFETCH](http://www.ebi.ac.uk/cgi-bin/emblfetch) Database accession no. AC022335 XP002188259**
- **HSIANG B ET AL: "A NOVEL HUMAN HEPATIC ORGANIC ANION TRANSPORTING POLYPEPTIDE (OATP2)" JOURNAL OF BIOLOGICAL CHEMISTRY, AMERICAN SOCIETY OF BIOLOGICAL CHEMISTS, BALTIMORE, MD, US, vol. 274, no. 52, 1999, pages 37161-37168, XP000877290 ISSN: 0021-9258**
- **ULBRECHT M ET AL.: "Assoziation of beta 2-adrenoreceptor variants with bronchial hyperresponsiveness" AMERICAN JOURNAL OF RESPIRATORY AND CRITICAL CARE MEDICINE, vol. 161, no. 2, February 2000 (2000-02), pages 469-474, XP002188594**

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**EP 1 186 672 B1**

## Description

[0001] This invention relates to polymorphisms in the human OATPC gene and corresponding novel allelic polypeptides encoded thereby. The invention also relates to methods and materials for analysing allelic variation in the OATPC gene, and to the use of OATPC polymorphism in treatment of diseases with OATPC transportable drugs.

[0002] Na<sup>+</sup>-independent organic anion transporting polypeptide (OATP) C gene is a member of the OATP supergene family involved in multifunctional transport of organic anion. OATPC transports the organic anion taurocholate, conjugated steroids: DHEAS, estradiol 17 $\beta$ -D-glucuronide and estrone-3-sulfate, eicosanoids: PGE<sub>2</sub>, thromboxane B<sub>2</sub>, leukotriene C<sub>4</sub>, and E<sub>4</sub>, and thyroid hormones T<sub>4</sub> and T<sub>3</sub><sup>1,2</sup>. OATPC has also been shown to be involved in the transport of xenobiotics, and drugs involved in lipid lowering *e.g.* statins<sup>1</sup>. Statins have been referred to as a first-line therapy for patients with atherosclerotic vascular diseases. The OATPC gene and its product is also thought to be of importance in other diseases due to its transport of DHEAS an adrenal steroid which has been suggested to have positive neuropsychiatric, immune, and metabolic effects<sup>3</sup>. Due to the substrate specificity, location in the liver, and being exclusively expressed in the liver, Abe *et al* suggested that OATPC could be the predominant clearance mechanism of several endogenous and exogenous substrates in the liver. OATPC is the first human molecule reported to transport thyroid hormones<sup>2</sup>.

[0003] This liver specific transporter may be useful in liver-specific drug delivery systems and liver-specific chemotherapy, bile acid formation and the pathogenesis of diseases such as cholestasis, hyperbilirubinemia and thyroid hormone resistance.

[0004] The OATPC gene (sometimes called OAPT2 in the literature) has been cloned by four different groups, annotated and published as EMBL accession numbers AB026257 (OATPC, 2452bp), AF205071 (OATP2, 2830, ref 1), AJ132573 (OATP2, 2778)<sup>4</sup> and AF060500 (LST-1)<sup>2</sup>. Polymorphism has been reported by Tamai<sup>5</sup> which is Asn 130Asp and Val174Ala although any functional effect was stated therein to be not clear. König (2000) J Biol Chem 275, 23161-23168 describes the genomic organisation of OATP 1, 2 and 8. International patent application WO 00/08157 describes human anion transporter genes and some polymorphisms.

[0005] All positions herein of polymorphisms in the OATPC polynucleotide relate to the position in one of SEQ ID NO 1 or 3-12 unless stated otherwise or apparent from the context.

[0006] All positions herein of polymorphisms in the OATPC polypeptide relate to the position in SEQ ID NO 2 unless stated otherwise or apparent from the context.

[0007] One approach is to use knowledge of polymorphisms to help identify patients most suited to therapy with particular pharmaceutical agents (this is often termed "pharmacogenetics"). Pharmacogenetics can also be used in pharmaceutical research to assist the drug selection process. Polymorphisms are used in mapping the human genome and to elucidate the genetic component of diseases. The reader is directed to the following references for background details on pharmacogenetics and other uses of polymorphism detection: Linder *et al.* (1997), Clinical Chemistry, **43**, 254; Marshall (1997), Nature Biotechnology, **15**, 1249; International Patent Application WO 97/40462, Spectra Biomedical; and Schafer *et al.* (1998), Nature Biotechnology, **16**, 33.

[0008] Clinical trials have shown that patient response to treatment with pharmaceuticals is often heterogeneous. Thus there is a need for improved approaches to pharmaceutical agent design and therapy.

[0009] Point mutations in polypeptides will be referred to as follows: natural amino acid (using 1 or 3 letter nomenclature), position, new amino acid. For (a hypothetical) example "D25K" or "Asp25Lys" means that at position 25 an aspartic acid (D) has been changed to lysine (K). Multiple mutations in one polypeptide will be shown between square brackets with individual mutations separated by commas.

[0010] The present invention is based on the discovery of polymorphisms in OATPC.

[0011] According to one aspect of the present invention there is provided a method for the detection of a polymorphism in OATPC in a human, which method comprises determining the sequence of the human at:

position 1561 of the OATPC gene as defined by the position in SEQ ID NO: 1 is not G; or  
position 488 in OATPC polypeptide defined by position in SEQ ID NO: 2 is not Gly.

[0012] The term human includes both a human having or suspected of having a OATPC mediated disease and an asymptomatic human who may be tested for predisposition or susceptibility to such disease. At each position the human may be homozygous for an allele or the human may be a heterozygote.

<sup>2</sup> Identification of a Novel Gene Family Encoding Human liver-specific Organic Anion Transporter LST-1, Takaaki Abe *et al* J Biol Chem **274**, 17159-17163 (1999)

<sup>1</sup> A Novel Human Hepatic Organic Anion Transporting Polypeptide (OATP2), Hsiang *et al* J Biol Chem **274**, 37161-37168 (1999)

<sup>3</sup> Bates *et al* (1998) Curr. Opin. Endocrinol. Diab. **5**, 357-366

<sup>4</sup> A novel human organic anion transporting polypeptide localised to the basolateral hepatocyte membrane, König Jorg *et al* (2000) Am J Physiol. Gastrointest. Liver Physiol. **278**: G156-G164

<sup>5</sup> Tamai *et al* (2000), BBRC, 273, 251-60

[0013] The term polymorphism includes single nucleotide substitution, nucleotide insertion and nucleotide deletion which in the case of insertion and deletion includes insertion or deletion of one or more nucleotides at a position of a gene.

[0014] The method for diagnosis is preferably one in which the sequence is determined by a method selected from amplification refractory mutation system and restriction fragment length polymorphism.

[0015] The status of the individual may be determined by reference to allelic variation at any one, two, three, four, five, six, seven, eight, nine or more positions.

[0016] The test sample of nucleic acid is conveniently a sample of blood, bronchoalveolar lavage fluid, sputum, or other body fluid or tissue obtained from an individual. It will be appreciated that the test sample may equally be a nucleic acid sequence corresponding to the sequence in the test sample, that is to say that all or a part of the region in the sample nucleic acid may firstly be amplified using any convenient technique e.g. PCR, before analysis of allelic variation.

[0017] It will be apparent to the person skilled in the art that there are a large number of analytical procedures which may be used to detect the presence or absence of variant nucleotides at one or more polymorphic positions of the invention. In general, the detection of allelic variation requires a mutation discrimination technique, optionally an amplification reaction and optionally a signal generation system. Table 1 lists a number of mutation detection techniques, some based on the PCR. These may be used in combination with a number of signal generation systems, a selection of which is listed in Table 2. Further amplification techniques are listed in Table 3. Many current methods for the detection of allelic variation are reviewed by Nollau *et al.*, Clin. Chem. **43**, 1114-1120, 1997; and in standard textbooks, for example "Laboratory Protocols for Mutation Detection", Ed. by U. Landegren, Oxford University Press, 1996 and "PCR", 2<sup>nd</sup> Edition by Newton & Graham, BIOS Scientific Publishers Limited, 1997.

#### Abbreviations:

[0018]

ALEX™	Amplification refractory mutation system linear extension
APEX	Arrayed primer extension
ARMS™	Amplification refractory mutation system
b-DNA	Branched DNA
bp	base pair
CMC	Chemical mismatch cleavage
COPS	Competitive oligonucleotide priming system
DGGE	Denaturing gradient gel electrophoresis
FRET	Fluorescence resonance energy transfer
HMG-CoA	3-hydroxy-3-methylglutaryl-coenzyme A
LCR	Ligase chain reaction
MASDA	Multiple allele specific diagnostic assay
NASBA	Nucleic acid sequence based amplification
OATP	Na <sup>+</sup> -independent organic anion transporting polypeptide
OLA	Oligonucleotide ligation assay
PCR	Polymerase chain reaction
PTT	Protein truncation test
RFLP	Restriction fragment length polymorphism
SDA	Strand displacement amplification
SNP	Single nucleotide polymorphism
SSCP	Single-strand conformation polymorphism analysis
SSR	Self sustained replication

(continued)

TGGE	Temperature gradient gel electrophoresis
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Table 1 - Mutation Detection Techniques

**[0019] General:** DNA sequencing, Sequencing by hybridisation

**Scanning:** PTT\*, SSCP, DGGE, TGGE, Cleavase, Heteroduplex analysis, CMC, Enzymatic mismatch cleavage

\* Note: not useful for detection of promoter polymorphisms.

#### Hybridisation Based

**[0020]** Solid phase hybridisation: Dot blots, MASDA, Reverse dot blots, Oligonucleotide arrays (DNA Chips)

Solution phase hybridisation: Taqman™ - US-5210015 & US-5487972 (Hoffmann-La Roche), Molecular Beacons - Tyagi *et al* (1996), Nature Biotechnology, 14, 303; WO 95/13399 (Public Health Inst., New York)

**Extension Based:** ARMS™, ALEX™ - European Patent No. EP 332435 B1 (Zeneca Limited), COPS - Gibbs *et al* (1989), Nucleic Acids Research, 17, 2347.

**Incorporation Based:** Mini-sequencing, APEX

**Restriction Enzyme Based:** RFLP, Restriction site generating PCR

**Ligation Based:** OLA

**Other:** Invader assay

Table 2 - Signal Generation or Detection Systems

**[0021] Fluorescence:** FRET, Fluorescence quenching, Fluorescence polarisation - United Kingdom Patent No. 2228998 (Zeneca Limited)

**Other:** Chemiluminescence, Electrochemiluminescence, Raman, Radioactivity, Colorimetric, Hybridisation protection assay, Mass spectrometry

Table 3 - Further Amplification Methods

**[0022]** SSR, NASBA, LCR, SDA, b-DNA

**[0023]** Preferred mutation detection techniques include ARMS™, ALEX™, COPS, Taqman, Molecular Beacons, RFLP, and restriction site based PCR and FRET techniques.

**[0024]** Particularly preferred methods include ARMS™ and RFLP based methods. ARMS™ is an especially preferred method.

**[0025]** In a further aspect, the diagnostic methods of the invention are used to assess the pharmacogenetics of a drug transportable by OATPC.

**[0026]** Assays, for example reporter-based assays, may be devised to detect whether one or more of the above polymorphisms affect transcription levels and/or message stability.

**[0027]** Individuals who carry particular allelic variants of the OATPC gene may therefore exhibit differences in their ability to regulate protein biosynthesis under different physiological conditions and will display altered abilities to react to different diseases. In addition, differences arising as a result of allelic variation may have a direct effect on the response of an individual to drug therapy. The diagnostic methods of the invention may be useful both to predict the clinical response to such agents and to determine therapeutic dose.

**[0028]** In a further aspect, the diagnostic methods of the invention, are used to assess the predisposition and/or susceptibility of an individual to diseases mediated by OATPC. This may be particularly relevant in the development of hyperlipoproteinemia and cardiovascular disease and the present invention may be used to recognise individuals who are particularly at risk from developing these conditions.

**[0029]** In a further aspect, the diagnostic methods of the invention are used in the development of new drug therapies which selectively target one or more allelic variants of the OATPC gene. Identification of a link between a particular allelic variant and predisposition to disease development or response to drug therapy may have a significant impact on the design of new drugs. Drugs may be designed to regulate the biological activity of variants implicated in the disease process whilst minimising effects on other variants.

**[0030]** In a further diagnostic aspect of the invention the presence or absence of variant nucleotides is detected by reference to the loss or gain of, optionally engineered, sites recognised by restriction enzymes.

[0031] According to another aspect of the present invention there is provided a human OATPC gene or its complementary strand comprising a variant allelic polymorphism at one or more of positions defined herein or a fragment thereof of at least 20 bases comprising at least one novel polymorphism.

[0032] Fragments are at least 17 bases, more preferably at least 20 bases, more preferably at least 30 bases.

[0033] According to another aspect of the present invention there is provided a polynucleotide comprising at least 20 bases of the human OATPC gene and comprising an allelic variant selected from any one of the following:

Region	variant	Position in SEQ ID NO	SEQ ID NO
Exon 10	C	1561	1

[0034] According to another aspect of the present invention there is provided a human OATPC gene or its complementary strand comprising a polymorphism, preferably corresponding with one or more the positions defined herein or a fragment thereof of at least 20 bases comprising at least one polymorphism.

[0035] Fragments are at least 17 bases, more preferably at least 20 bases, more preferably at least 30 bases.

[0036] The invention further provides a nucleotide primer which can detect a polymorphism of the invention.

[0037] According to another aspect of the present invention there is provided an ARMS allele specific primer or an allele-specific oligonucleotide probe selected from one of the following:

an ARMS allele specific primer;  
or an allele-specific oligonucleotide probe;  
capable of detecting a OATPC gene polymorphism, preferably at one or more of the positions as defined herein.

[0038] An allele specific primer is used, generally together with a constant primer, in an amplification reaction such as a PCR reaction, which provides the discrimination between alleles through selective amplification of one allele at a particular sequence position e.g. as used for ARMS™ assays. The allele specific primer is preferably 17- 50 nucleotides, more preferably about 17-35 nucleotides, more preferably about 17-30 nucleotides.

[0039] An allele specific primer preferably corresponds exactly with the allele to be detected but derivatives thereof are also contemplated wherein about 6-8 of the nucleotides at the 3' terminus correspond with the allele to be detected and wherein up to 10, such as up to 8, 6, 4, 2, or 1 of the remaining nucleotides may be varied without significantly affecting the properties of the primer.

[0040] Primers may be manufactured using any convenient method of synthesis. Examples of such methods may be found in standard textbooks, for example "Protocols for Oligonucleotides and Analogues; Synthesis and Properties," Methods in Molecular Biology Series; Volume 20; Ed. Sudhir Agrawal, Humana ISBN: 0-89603-247-7; 1993; 1<sup>st</sup> Edition. If required the primer(s) may be labelled to facilitate detection.

[0041] According to another aspect of the present invention there is provided an allele-specific oligonucleotide probe capable of detecting a OATPC gene polymorphism, preferably at one or more of the positions defined herein.

[0042] The allele-specific oligonucleotide probe is preferably 17- 50 nucleotides, more preferably about 17-35 nucleotides.

[0043] The design of such probes will be apparent to the molecular biologist of ordinary skill. Such probes are of any convenient length such as up to 50 bases, up to 40 bases, more conveniently up to 30 bases in length, such as for example 8-25 or 8-15 bases in length. In general such probes will comprise base sequences entirely complementary to the corresponding wild type or variant locus in the gene. However, if required one or more mismatches may be introduced, provided that the discriminatory power of the oligonucleotide probe is not unduly affected. The probes of the invention may carry one or more labels to facilitate detection.

[0044] According to another aspect of the present invention there is provided an allele specific primer or an allele specific oligonucleotide probe capable of detecting a OATPC gene polymorphism at one of the positions defined herein.

[0045] In another aspect of the invention, the single nucleotide polymorphisms of this invention may be used as genetic markers in linkage studies. This particularly applies to the polymorphisms of relatively high frequency. The OATPC gene is on chromosome 12p (as shown from a database search with the cDNA as a query sequence). Low frequency polymorphisms may be particularly useful for haplotyping as described below. A haplotype is a set of alleles found at linked polymorphic sites (such as within a gene) on a single (paternal or maternal) chromosome. If recombination within the gene is random, there may be as many as 2<sup>n</sup> haplotypes, where 2 is the number of alleles at each SNP and n is the number of SNPs. One approach to identifying mutations or polymorphisms which are correlated with clinical response is to carry out an association study using all the haplotypes that can be identified in the population of interest. The frequency of each haplotype is limited by the frequency of its rarest allele, so that SNPs with low frequency alleles are particularly useful as markers of low frequency haplotypes. As particular mutations or polymorphisms associated with certain clinical features, such as adverse or abnormal events, are likely to be of low frequency

within the population, low frequency SNPs may be particularly useful in identifying these mutations (for examples see: Linkage disequilibrium at the cystathionine beta synthase (CBS) locus and the association between genetic variation at the CBS locus and plasma levels of homocysteine. *Ann Hum Genet* (1998) **62**:481-90, De Stefano V, Dekou V, Nicaud V, Chasse JF, London J, Stansbie D, Humphries SE, and Gudnason V; and Variation at the von willebrand factor (vWF) gene locus is associated with plasma vWF:Ag levels: identification of three novel single nucleotide polymorphisms in the vWF gene promoter. *Blood* (1999) **93**:4277-83, Keightley AM, Lam YM, Brady JN, Cameron CL, Lillicrap D).

**[0046]** According to another aspect of the present invention there is provided use of a statin in preparation of a medicament for treating a cardiovascular disease in a human detected as having an OATPC polymorphism at one or more of the following positions:

position 1561 of the OATPC gene as defined by the position in SEQ ID NO: 1 is not G;  
position 488 in OATPC polypeptide defined by position in SEQ ID NO: 2 is not Gly.

**[0047]** Preferably determination of the status of the human is clinically useful. Examples of clinical usefulness include deciding which statin drug or drugs to administer and/or in deciding on the effective amount of the statin drug or drugs. Statins already approved for use in humans include atorvastatin, cerivastatin, fluvastatin, pravastatin and simvastatin. The reader is referred to the following references for further information: Drugs and Therapy Perspectives (12<sup>th</sup> May 1997), **9**: 1-6; Chong (1997) Pharmacotherapy **17**: 1157-1177; Kellick (1997) Formulary **32**: 352; Kathawala (1991) Medicinal Research Reviews, **11**: 121-146; Jahng (1995) Drugs of the Future **20**: 387-404, and Current Opinion in Lipidology, (1997), **8**, 362 - 368. A preferred statin drug is compound 3a (S-4522) in Watanabe (1997) Bioorganic and Medicinal Chemistry **5**: 437-444; now called rosuvastatin, see Olsson (2001) American Journal of Cardiology, **87**, supplement 1, 33-36. The term "drug transportable by OATPC" means that transport by OATPC in humans is an important part of a drug exerting its pharmaceutical effect in man. For example, some statins have to be transported to the liver by OATPC to exert their lipid lowering effects.

**[0048]** According to another aspect of the present invention there is provided an allelic variant of human OATPC polypeptide comprising:

an arginine at position 488 of SEQ ID NO 2;  
or a fragment thereof comprising at least 10 amino acids provided that the fragment comprises at least one allelic variant.

**[0049]** Fragments of polypeptide are at least 10 amino acids, more preferably at least 15 amino acids, more preferably at least 20 amino acids.

**[0050]** The invention will now be illustrated but not limited by reference to the following Examples. All temperatures are in degrees Celsius.

**[0051]** In the Examples below, unless otherwise stated, the following methodology and materials have been applied.

**[0052]** AMPLITAQ™, available from Perkin-Elmer Cetus, is used as the source of thermostable DNA polymerase.

**[0053]** General molecular biology procedures can be followed from any of the methods described in "Molecular Cloning - A Laboratory Manual" Second Edition, Sambrook, Fritsch and Maniatis (Cold Spring Harbor Laboratory, 1989).

**[0054]** Electropherograms were obtained in a standard manner: data was collected by ABI377 data collection software and the wave form generated by ABI Prism sequencing analysis (2.1.2).

## Example 1

### **Identification of Polymorphisms**

#### **1. Methods**

##### DNA Preparation

**[0055]** DNA was prepared from frozen blood samples collected in EDTA following protocol I (Molecular Cloning: A Laboratory Manual, p392, Sambrook, Fritsch and Maniatis, 2<sup>nd</sup> Edition, Cold Spring Harbor Press, 1989) with the following modifications. The thawed blood was diluted in an equal volume of standard saline citrate instead of phosphate buffered saline to remove lysed red blood cells. Samples were extracted with phenol, then phenol/chloroform and then chloroform rather than with three phenol extractions. The DNA was dissolved in deionised water.

Template Preparation

[0056] Templates were prepared by PCR using the oligonucleotide primers and annealing temperatures set out below. The extension temperature was 72° and denaturation temperature 94°. Generally 50 ng of genomic DNA was used in each reaction and subjected to 35 cycles of PCR. Where described below, the primary fragment was diluted 1/100 and two microlitres were used as template for amplification of secondary fragments. PCR was performed in two stages (primary fragment then secondary fragment) to ensure specific amplification of the desired target sequence.

Polymorphisms in OATPC: cDNA screening of 15 Liver samples				
Region	SNP	Position	Amino Acid Change	Allele frequencies
Exon 4	G/A	510	None	G=96.7% A=3.3%
Exon 5	C/T	670	None	C=50% T=50%
Exon 5	C/T	696	None	C=60% T=40%
Exon 9	C/G	1299	Phe400Leu	C=96.7% G=3.3%
Exon 9	G/A	1312	Va1405Ile	G=96.7% A=3.3%
Exon 9	G/A	1347	None	G=96.7% A=3.3%
Exon 10	G/C	1561	Gly488Arg	G=96.7% C=3.3%
Exon 14	A/C	2028	Leu643Phe	A=90% C=10%

[0057] OATP2 above refers to the clone sequenced by Hsiang et al (ref 1). Some comment on the numbering of exons in OATPC is required. This gene contains an exon (38 bp) upstream (5' UTR region) of the exon containing the ATG start site for translation. Therefore the exon numbering could vary depending whether this exon is counted as the first exon or not. In the literature, Konig (2000) JBC 275: 23161-68, have defined exon 1 as that containing the ATG start site and therefore we have adopted the same numbering in this application (but note that the priority document relating to the present application did vice versa; for example, exon 5 in this application is equivalent to exon 6 in the priority document).

PCR PRODUCTS		
Fragment	Forward Oligo	Reverse Oligo
443-999	443-466	979-999
874-1360	874-896	1337-1360
1255-1684	1255-1278	1663-1684
1559-2095	1559-1581	2073-2095

RFLP analysis			
Polymorphism	Position	RFLP Enzyme/PCR size	RLFP fragment size
G/A	510		

# EP 1 186 672 B1

(continued)

RFLP analysis			
Polymorphism	Position	RFLP Enzyme/PCR size	RFLP fragment size
C/T	670	BmR I/ 595bp	C=349bp, 246bp, T=595bp
C/T	696		
C/G	1299	Apo I/ 595bp	C=30bp, 60bp, 380 bp G=90bp, 380bp
G/A	1312	Bst 4Cl/ 595bp	G=77bp, 393bp A=470bp
G/A	1347		
G/C	1561	HpyCH4IV/ 595bp	G=470bp C=144bp, 326bp
A/C	2028	Ase I/ 595bp	A=89bp, 488bp C=577bp

Example 2

[0058]



<b>Further OATPC Polymorphisms</b>							
<b><u>SNPs in OATPC 3'UTR (positions according to SEQ ID NO 1)</u></b>							
<b>Exon</b>	<b>Nucleotide</b>	<b>SNP</b>				<b>Frequency</b>	<b>Frequency</b>
						<b>Caucasian</b>	<b>Japanese</b>
3' UTR	2327	Ins T				not screened	0.1
3' UTR	2342	T>C				not screened	0.4
<b><u>SNPs in OATPC promoter (positions according to SEQ ID NO 3)</u></b>							
	<b>Nucleotide</b>	<b>SNP</b>				<b>Frequency</b>	<b>Frequency</b>
						<b>Caucasian</b>	<b>Japanese</b>
	321	T>G				0.03	not screened
	1332	A>C				0.08	not screened
<b><u>SNPs in OATPC introns</u></b>							
<b>Intron</b>	<b>Nucleotide</b>	<b>SNP</b>	<b>Nucleotide</b>	<b>Sequence</b>			
	<b>position in</b>		<b>position</b>	<b>ID No</b>			
	<b>relation to</b>		<b>in</b>				
	<b>exon</b>		<b>sequence</b>				
1	IVS1+21	T>A	41	4			
2	IVS2+89	T>G	109	5			
2	IVS2+224	A>G	244	5			
3	IVS3+97	C>A	117	6			
3	IVS3+263	G>A	283	6			
4	IVS4+189	G>A	209	7			
4	IVS4+191	G>A	211	7			
4	IVS5-118	delCTTGTA	63	8			
6	IVS6+33	C>T	53	9			
9	IVS10-107	ins TTC	75	10			
11	IVS11+142	Ins T	162	11			
12	IVS13-97	G>C	84	12			

**OATPC intronic SNPs****Key**

[0059] 20bp of exon sequence shown in uppercase

# EP 1 186 672 B1

Intron sequence in lowercase (200 to 300bp only)  
SNP shown in uppercase (one allele only)

5 **Sequence ID No 4**  
IVS1+21 T>A SNP at position 41 in this sequence

GATACTGCAA	TGGATTGAAG	gtagaataag	ttttatgttt	Ttgagctaaa	ataagtaa	60
agggaaacttt	aatgtataga	aaagcaagtt	gttaaaaaaga	acattatgtt	tcaaattata	120
attttcaatt	gaagcatata	ttgaaatatt	aacataatga	ttcatacctt	gattttaa	180
agtccttttaa	tctgattaag	200				

15 **Sequence ID No 5**  
IVS2+89 T>G SNP at position 109 in this sequence  
IVS2+224 A>G SNP at position 244 in this sequence

TGACGGAAGC	TTTGAAATTG	gtaacatttta	ttttctattt	taataaccaa	acttgcaaag	60
ttaaaaaata	tatatgcttt	acaccactgg	ttatcaactg	gggtaaatTt	atctctcaca	120
ggcaatttgg	caataactaa	aaacatttgt	ggttgtcata	actgcacagg	ggttgggggc	180
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50 **Sequence ID No 8**  
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Sequence ID No 9 IVS6+33 C>T SNP at position 53 in this sequence  
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Sequence ID No 10  
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AAATAATCCA GTGACATCTC 200

Sequence ID No 11  
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#### OATPC promoter region

35 [0060] Total length of the sequence = 1538bp  
1500bp of OATPC sequence directly upstream from the cDNA sequence  
Sequence in uppercase represents 38bp overlap with the cDNA sequence (SEQ ID NO 1) where this 38bp is 5'UTR  
sequence.  
Nucleotide positions in the promoter have been determined where the -1 position is the base (lowercase) directly  
40 upstream of the end of the cDNA sequence.

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EP 1 186 672 B1

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EP 1 186 672 B1

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EP 1 186 672 B1

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# EP 1 186 672 B1

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       tgcttaatat   aattagaaag   ttacaagtag   gaaataaatg   tattactaat   cagaataaat   180  
       ataaaatcca   gtccttattt   200

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<400> 10

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cccttctttt gtctttttct tctctctctc tctttttgat atatgtctat catatatttc 180
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ggctgtaatt aataattttg c                                     201

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<210> 12  
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<400> 12

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tcattgcagtt acatttaaaa tatgttcctt aaactgacat cttctcttct cctattacag 180
gaggaattct agctccaata,                                     200

```

## Claims

1. A method for the detection of a polymorphism in OATPC in a human, which method comprises determining the sequence of the human at:

position 1561 of the OATPC gene as defined by the position in SEQ ID NO: 1 is not G; or  
 position 488 in OATPC polypeptide defined by position in SEQ ID NO: 2 is not Gly.

2. Use of a method as defined in claim 1 to assess the pharmacogenetics of a drug transportable by OATPC.
3. A polynucleotide comprising at least 20 bases of the human OATPC gene and comprising the allelic variant:

Region	variant	Position in SEQ ID NO	SEQ ID NO
Exon 10	C	1561	1

4. An ARMS allele specific primer or an allele-specific oligonucleotide probe selected from one of the following:

an ARMS allele specific primer; or  
an allele-specific oligonucleotide probe,  
capable of detecting a OATPC gene polymorphism as defined in claim 1.

5. Use of an OATPC polymorphism as defined in claim 1 as a genetic marker in a linkage study.

6. Use of a statin in preparation of a medicament for treating a cardiovascular disease in a human detected as having an OATPC polymorphism at one or more of the following positions:

position 1561 of the OATPC gene as defined by the position in SEQ ID NO: 1 is not G;  
position 488 in OATPC polypeptide defined by position in SEQ ID NO: 2 is not Gly.

7. A use according to claim 6 in which the drug is rosuvastatin.

8. An allelic variant of human OATPC polypeptide comprising;  
an arginine at position 488 of SEQ ID NO 2  
or a fragment thereof comprising at least 10 amino acids provided that the fragment comprises the allelic variant.

### Patentansprüche

1. Verfahren zum Nachweis eines Polymorphismus in OATPC in einem Menschen, wobei man in dem Verfahren bestimmt, daß die Sequenz des Menschen in:

Position 1551 des OATPC-Gens, wie durch die Position in SEQ ID NO: 1 definiert, nicht G ist; oder  
Position 488 im OATPC-Polypeptid, definiert durch die Positionen in SEQ ID NO: 2, nicht Gly ist.

2. Verwendung eines Verfahrens gemäß Anspruch 1 zur Beurteilung der Pharmacogenetik eines durch OATPC transportierbaren Arzneistoffs.

3. Polynukleotid, umfassend wenigstens 20 Basen des menschlichen OATPC-Gens und umfassend die Allelvariante:

Region	Variante	Position in SEQ ID NO:	SEQ ID NO:
Exon 10	C	1561	1

4. ARMS-Allel-spezifischer Primer oder Allelspezifische Oligonukleotidsonde, ausgewählt aus einem der folgenden Moleküle:

einem ARMS-Allel-spezifischen Primer; oder  
einer Allel-spezifischen Oligonukleotidsonde,  
womit ein OATPC-Gen-Polymorphismus gemäß Anspruch 1 nachgewiesen werden kann.

5. Verwendung eines OATPC-Polymorphismus gemäß Anspruch 1 als genetischer Marker in einer Verknüpfungsuntersuchung.

6. Verwendung eines Statins bei der Herstellung eines Arzneimittels zur Behandlung einer Herz-Kreislauf-erkrankung in einem Menschen, bei dem ein OATPC-Polymorphismus an einer oder mehreren der folgenden Positionen nachgewiesen wurde:

Position 1561 des OATPC-Gens, wie durch die Positionen in SEQ ID NO: 1 definiert, ist nicht G;  
Position 488 im OATPC-Polypeptid, definiert durch die Positionen in SEQ ID NO: 2, ist nicht Gly.

7. Verwendung nach Anspruch 6, wobei es sich bei dem Arzneistoff um Rosuvastatin handelt.

8. Allelvariante des menschlichen OATPC-Polypeptids, umfassend:

ein Arginin in Position 488 der SEQ ID NO: 2  
oder ein Fragment davon, umfassend wenigstens 10 Aminosäuren, vorausgesetzt, daß das Fragment die Allelvariante umfaßt.

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## Revendications

1. Procédé pour la détection d'un polymorphisme dans un OATPC d'un être humain, lequel procédé comprend l'étape consistant à déterminer la séquence de l'humain au niveau de :

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la position 1561 du gène d'OATPC, comme cela est défini par la position dans la SEQ ID n° : 1, qui n'est pas un G ; ou  
la position 488 dans un polypeptide d'OATPC, définie par la position dans la SEQ ID n° : 2, qui n'est pas une Gly.

2. Utilisation d'un procédé, comme cela est défini à la revendication 1, pour évaluer les aspects pharmacogénétiques d'une substance médicamenteuse pouvant être transportée par un OATPC.

3. Polynucléotide comprenant au moins 20 bases du gène d'OATPC humain et comprenant le variant allélique :

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Région	Variant	Position dans la SEQ ID n° :	SEQ ID n° :
Exon 10	C	1561	1

4. Amorce spécifique d'un allèle ARMS ou sonde oligonucléotidique spécifique d'un allèle choisie parmi l'un des éléments suivants :

une amorce spécifique d'un allèle ARMS ; ou  
une sonde oligonucléotidique spécifique d'un allèle,  
capable de détecter un polymorphisme dans un gène d'OATPC, comme cela est défini à la revendication 1.

5. Utilisation d'un polymorphisme d'OATPC, comme cela est défini à la revendication 1, en tant que marqueur génétique dans une étude de liaison.

6. Utilisation d'une statine dans la préparation d'un médicament destiné à traiter une affection cardiovasculaire chez un être humain, chez qui on a détecté un polymorphisme d'OATCP au niveau d'une ou plusieurs positions parmi les suivantes :

la position 1561 du gène d'OATPC, comme cela est défini par la position dans la SEQ ID n° : 1, n'est pas un G ; ou  
la position 488 dans un polypeptide d'OATPC, définie par la position dans la SEQ ID n° : 2, n'est pas une Gly.

7. Utilisation selon la revendication 6, dans laquelle la substance médicamenteuse est la rosuvastatine.

8. Variant allélique d'un polypeptide d'OATPC humain comprenant :

une arginine à la position 488 de la SEQ ID n° : 2 ou un fragment de celle-ci comprenant au moins 10 acides aminés, à condition que le fragment comprenne le variant allélique.

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